Linzer biol. Beitr.	33/1	103-136	29.6.2001

On the Staphylinidae of Greece. II. New species and new records from central and northern Greece (Insecta: Coleoptera)

V. ASSING & P. WUNDERLE

A b s t r a c t: The results of a field trip to central and northern Greece in spring 2000 are presented. 188 identified species of Staphylinidae are reported, among them several new taxa and first records from Greece. 10 species are described, illustrated, and distinguished from similar congeners: Deliphrosoma angulatum sp. n. (Fokis, Fthiotis), Pareudectus vitsiensis sp. n. (Florina), Sunius fokisensis sp. n. (Fokis), Lathrobium vorasensis sp. n. (Florina), Tectusa rastrifera sp. n. (Fokis, Fthiotis), T. longiuter sp. n. (Evritania), T. viduus sp. n. (Evritania), T. timfristosensis sp. n. (Evritania), T. vardousiensis sp. n. (Fokis), and T. vrontousensis sp. n. (Séres). Anthobium ganglbaueri (LUZE), Quedius bernhaueri RAMBOUSEK, Leptusa jelineki PACE, L. peristerica PACE, L. winneguthiana PACE, and L. sororella PACE are for the first time recorded from Greece. The facies and male primary sexual characters of Q. bernhaueri are illustrated.

K e y w o r d s: Coleoptera, Staphylinidae, Palaearctic region, Greece, taxonomy, biogeography, new species, new records

Introduction

In the past, the staphylinid fauna of Greece has largely been neglected in comparison to many other parts of Europe. Comprehensive taxonomic works, keys, and species catalogues focussing on this region are not available. All that is currently known about Greek Staphylinidae is contained in some older taxonomic works (e. g. EPPELSHEIM 1888, KRAATZ 1858a, 1858b), some comprehensive systematic and taxonomic synopses covering larger areas (e. g. ASSING 1997, COIFFAIT 1972, 1974, 1978, 1982, 1984, PACE 1989, PUTHZ 1971, ZERCHE 1990, in prep.), recent revisions of individual genera from Greece (e. g. ASSING 1999, 2000), various shorter taxonomic papers (usually individual species desriptions, and several, mostly older species lists summarizing the results of individual field trips (e. g. SCHEERPELTZ 1931, 1956, 1957, 1959, 1963, 1964). More recent studies as well as revisions currently in progress show that our knowledge especially of the more or less endemic montane and alpine fauna of the numerous Greek mountain ranges is far from complete (ASSING 1999, 2000, ASSING & WUNDERLE 1999, ZERCHE in prep.). However, there are also many other, more widespread Staphylinidae (e. g. Proteinus, some Tachyporinae, many Aleocharinae, to mention only a few) which require thorough taxonomic and biogeographic treatment.

In recent years, numerous field trips designed to study the staphylinid fauna of various parts of Greece have been organized by the authors and several colleagues. Below, we present the results of one such excursion to central and northern Greece in spring 2000.

104

Measurements and abbreviations

The measurem	nents are given in mm and abbreviated as follows:
AL	length of antennae
HL	head length from anterior margin of clypeus to hind margin
HW	.head width
PL	length of pronotum along median line
PW	.width of of pronotum
EL	length of elytra at suture, from apex of scutellum to elytral hind margin
EW	.width of elytra
HTi	length of metatibia
НТа	length of metatarsi
ML	length of aedeagus (median lobe) from base to apex of ventral process
	total length of body (if not indicated otherwise).

The material listed and referred to below is deposited in the authors' collections (cAss, cWun), if not stated otherwise.

Results

In all, 2885 adult Staphylinidae of more than 214 species were recorded, among them several new taxa and first records from Greece (Tab. 1). The species and specimens were not named or identified when $\delta \delta$ were not available, but would have been necessary for a safe identification (e. g. *Xantholinus*, *Gabrius*, *Quedius*, *Mycetoporus*), in the case of genus group taxa which are in a state of taxonomic confusion (e. g. *Hydrosmecta*, *Mocyta*) and in which identification on the species level would require a comprehensive revision of the types, or when there was reasonable doubt regarding the identity of a species for other reasons (e. g. ambiguously or undescribed species).

Tab. 1: Staphylinidae recorded in central and northern Greece in spring 2000.

Localities: 1 – Evritania, 75km W Lamia, Timfristos, skiresort, 38°57N, 21°48E, 1900m, sifted grass, moss, etc. near snow, 14.IV.; 2 – Evritania, 10km S Karpenisi, 38°50′00N, 21°47′16E, 1230m, Abies forest in northern exposition, 14.IV.; 3 – Evritania, 15km SSW Karpenisi, Mt. Kaliakouda, 38°48′12N, 21°46′08E, 1500m, sifted grass, moss, litter of shrubs, etc. near snow, 14.IV.; 4 – Evritania, 13km SSW Karpenisi, Mt. Kaliakouda, 38°49′00N, 21°46′40E, 1200m, Abies forest, 14.IV.; 5 – Fthiotis, 30km SW Lamia, NE Amfiklia, 38°46′00N, 22°32′55E, bank of Kifisos river, 300m, 15.IV.; 6 – Fthiotis, Parnassos, Kelaria ski resort, 38°33′02N, 22°34′48E, 1780m, N-slope, sifted grass, moss, and Juniperus litter near snow, 15.IV.; 7 – Fthiotis, Parnassos, below Kelaria ski resort, 38°33 - 38°34N, 22°34E, 1730-1760m, Abies forests (Leptobium illyricum found under stone on pasture), 15.IV.; 8 – Fthiotis, 30km W Lamia, W Kalithea, 38°53N, 22°05E, 800m, Quercus and Abies forests, 16.IV.; 9 – Fthiotis, 30km W Lamia, bank of Sperchios river, 38°55′46N, 21°59′19E, river bank and riparian woodland, 16.IV.; 11 – Fokis, 45 km SSW Lamia, Oros Giona, W Vraila peak, 38°39′36N, 22°19′32E, 1700m, sifted grass, moss, etc. near snow, 17.IV.; 12 – Fokis, 45 km SSW Lamia, Oros Giona, 38°39′58N, 22°19′12E, 1650m, Abies forest in northern exposition, 17.IV.; 13 – Fokis, 45km SW Lamia, Vardousia Oros, 38°42′06N, 22°08′54E, 1600m, alpine meadow, sifted grass and shrub litter, partly near snow, 18.IV.; 14 – Fokis, 45km SW Lamia, Vardousia Oros, 38°41′37N, 22°09′47E, 1540m, Abies forest in northern exposition, 18.IV.; 15 – Fokis, ca. 35km SW Lamia, N Sikia, 38°41′23N, 22°12′54E, 745m, river bank, 18.IV.; 15 – Fokis, ca. 35km SW Lamia, N Sikia, 38°41′23N, 22°12′54E, 745m, river bank, 18.IV.; 16 – Florina, 15km S Florina, Oros Vitsi, 40°39′N, 21°23E, NW-slope of peak, 1900m, sifted grass, moss, etc., 21./25.IV.; 17 – Florina, 15km S Florina, Oros Vitsi, 40°39′N, 21°23E, NW-slope of peak, 1800-1880m,

beechwood with snow, sifted deep leaf litter and nest of Formica pratensis, 21./25.IV.; 18 – Florina, 15km S Florina, Oros Vitsi, 40°39'N, 21°23E, NW-slope of peak, 1800m, montane meadow, under stones, 21.IV.; 19 – Pela, 40km NE Florina, Oros Voras, 40°54'15N, 21°49'30E, 1950m, meadow, sifted grass and under stones near snow, 22.IV.; 20 – Florina, 40km NE Florina, Oros Voras, 40°52'58N, 21°47'36E, 1650m, beechwood, 22.IV.; 21 – Florina, 15km W Florina, Oros Varnous, 40°47'51N, 21°14'42E, 1600m, beechwood, 23.IV.; 22 – Florina, 10km S Florina, Drosopigi, 40°42N, 21°28E, 850m, oak forest, 23.IV.

locality	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
Micropeplus latus HAMPE 1861		Г	Г	Г	Γ	Г		Γ	Γ					ī						1	П	П
M. cf. turcicus COIFFAIT 1957	Γ	Γ		Г		Г	Г	41			Г	Г		Г							П	П
M. ripicola KERSTENS 1964	Г					Г				1												П
Micropeplus sp.		П			Г			7	Г			П										П
Proteinus ovalis STEPHENS 1834					1		1				4									2		1
P. brachypterus (F. 1792)		19	2	1				9			3		1	4		1				i	1	8
P. atomarius ERICHSON 1840																	1					
Proteinus sp. 1				1							5					2	10					
Proteinus sp. 2																				2		
Metopsia similis ZERCHE 1998																						ı
Phyllodrepa palpalis LUZE 1906							L	L						1								
P. floralis (PAYK. 1789)								1														
Omalium rivulare (PAYK. 1789)																						2
O. rugatum MULS. & REY 1880		1							L					1		Ш				1		1
O. cinnamomeum KRAATZ 1858								2														Ш
O. excavatum Stephens 1834	1	L		L	L		1	L	L		2			L		L						Ш
Anthobium melanocephalum (ILLIGER 1794)	L			1	L	L		L	L					4			1			i		Ш
A. aff. atrocephalum (GYLL. 1827)	L			L	L	L	L	L	L			Ц		1		2	9			2	1	ı
A. ganglbaueri (LUZE 1905)	L			L	L	L	L	L	L	L		Ц				L						5
Amphichroum canaliculatum (ERICHSON 1840)	L	L		L	L	L	L	L	L			Ц		Ш			2				Ш	Ц
Lesteva longoelytrata (GZE 1777)	L	L		L	L	L	L	L	L			Ц		Ц		10			33		2	Ш
Deliphrosoma angulatum sp. n.	L			L	L	1	1	L	L		2	Ц	2	L		Ш						Ц
D. fratellum (ROTTENBG, 1874)	1	Ц		L	L	L	L	L		Ц		Ц		Ц		Ш	Ш	Ц	Ш			Ц
Boreaphilus velox (HEER 1839)	L	L		L.	L	1	4		Ľ	Ц		Ц	2	Ш							Ш	Ц
Coryphiodes sp. n.	L	L		L	L	L	L		L	Ц		Ц		Ц		1					Ш	Ц
Pareudectus vitsiensis sp. n.	L	L			L	L	L	L	L	Ц		Ц				4					Ц	Ц
Carpelimus subtilis (ER. 1839)		L		L	L	L	L	L	L	6		Ц		Ш							Ц	Ц
C. fuliginosus (GRAV. 1802)	L	L		L	1	L	L	L	L	Ц		Ц		Ц								Ц
C. rivularis (MOTSCH. 1860)	L	Ц	Ц	L	L	L	L	L	L	1	L	Ц		Ц		Ц	Ц	Ц		Ц	Ц	Ц
C. bilineatus (STEPHENS 1834)	L								L	3		Ц							Щ			Ш
C. corticinus (GRAV. 1806)	L	L		L	L	L	L	L		2		Ш							Ц		Ц	2
Ochthephilus angustior (BERNHAUER 1943)	L	L		L	L		L	L		2		Ц		Ц			Ш	Ш				Ц
O. rosenhaueri (KIESENW. 1850)	L	Ц	Ц	$ldsymbol{ld}}}}}}$	$oldsymbol{ol}}}}}}}}}}}}}}$	L	L	L	L	5	Ц	Ц		Ц		Ц		Ш	Ш	Ш	Ш	Ц
O. venustulus (ROSENH. 1856)	L	L	Ц	L	2	Ļ	L	L	L	4	Ш	Ц		Ц							Ш	Ш
Anotylus inustus (GRAV. 1806)	L	$oxed{oxed}$		L	3	L	L	1	3	26	7	Ц	1	Ц		Ц	Ш	Щ			Ш	Ц
A. nitidulus (GRAV. 1802)		Ц		Ш	Ш	L		L	Ц	Ц	Ц	Ц	1	Ц		Ш	1	Ц	Ш	Ш	Ш	Ц
Platystethus nitens (SAHLB.1832)				L	1	<u> </u>			L	6	Ш	Ш		Ш		L	Ш					

locality	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
Bledius fossor HEER 1839	Г	Г	Τ	Г	Ī	Π	Γ	Π	Т	18	Г	Г	Г	Τ	T	Π			T		Γ	П
B. cribricollis HEER 1839	Г		Г	Г	1	Г	Г	Г	Г						Г			П	Г		Г	П
Stenus flavipalpis THOMS. 1860	┢	Г	Г	Г	Ė	Г	Г	Г	Г	Г	П	Г	Г	┢	Г				Г	1	Г	П
S. glacialis HEER 1839	Г	Г	Г	Г	T	Г	Т	Г		П		ī	П		П		2		П	Ė	Г	П
S. subaeneus ER. 1840	Г	Г	Г	Т	Г	Г	Г	ī	Г		Г	Г		Γ	Г			П	П	П	Г	П
S. turbulentus BONDROIT 1912	Г					Г		Г		2			П	Г	Г		П				Г	П
S. ochropus Kiesenw. 1858	Г		4	1	Γ		Г		Г				2	Г							П	5
Paederidus rubrothoracicus (GOEZE 1777)		Г	Г	Г						2				Г	ı						П	П
Paederus schoenherri Czw. 1889									Г				_	Г				3			П	П
Leptobium illyricum (ER. 1839)							1	Г	Г													П
Rugilus dilutipes (REITT. 1884)	Г	1	Г		Г			3	2	9			Г		П		П		П	П	Г	8
Astenus cf. procerus (GRAV. 1806)	Г					Г						П	1		П						Г	П
Scopaeus gracilis (SPERK 1835)	Г					Г		Г		1	П	П			1						Г	П
S. laevigatus (GYLLENHAL 1827)	Γ								Г	2		П		Г	П		1				П	П
S. debilis HOCHHUTH 1851		Г								2											П	П
Medon brunneus (ER. 1839)	Г	9		1			3	97	1					16							П	17
M. fusculus (MANNERH. 1830)									1	П												П
Pseudomedon dido (SAULCY 1864)										1	П	П			П						П	П
P. obscurellus (ER. 1840)	П									1				П	П			╗			П	П
Sunius fokisensis sp. n.													27					╗			٦	П
Lobrathium rugipenne (HOCHHUTH 1851)	П	П			1					П			1					٦			П	П
Lathrobium sp. n.?																	1	╗		\neg	П	П
Lathrobium vorasensis sp. n.																	П	ヿ		2	٦	П
Cryptobium collare (REITT. 1884)										33			\neg		П		\sqcap	ヿ	П	コ	П	П
Leptacinus pusillus (STEPH.1833)														1			T	П		T	٦	
Gyrohypnus angustatus STEPHENS 1833																	╗	╗	5		П	
Xantholinus decorus Er. 1839								3										П		П	П	
X. laevigatus JACOBSON 1874																	I			4		
Xantholinus sp. Q Q											\neg	\neg	٦		П	T]	╗	4	Ī	T	٦	П
Othius laeviusculus STEPH. 1832								4			٦	П	T		٦		٦	٦			٦	П
O. punctulatus (GOEZE 1777)											T				I		12			2	٦	
O. lapidicola MÄRK.KSW. 1848				8						П	T	Ī			П		12	П		1	╗	
Atrecus affinis (PAYKULL 1789)										\Box	\Box	\Box		33			4			1	П	
Erichsonius rivularis (Ksw.1858)										5	П		1					П			\exists	
Neobisnius prolixus (ER. 1840)													\Box		1				I		I	\Box
Philonthus rufimanus ER. 1840		╗			7	I	П	I		5			丁	T	T		I			П	\Box	
P. fumarius (GRAV. 1806)										4			T	T		П	1				П	
P. quisquiliarius (GYLL. 1810)		_]		_]	2	\Box	J	J	_]	J	J]	_]	J			_]			J	
P. rubripennis Stephens 1832				\Box				\Box		1			_]	\Box	J		J			⅃	J	
Gabrius toxotes JOY 1913					7	_]	_]	_]	_1			J					Ţ		3	_T	T	\Box
G. obenbergeri Smetana 1952						_]	_]	1						_	_]]	3
G. ravasinii GRIDELLI 1920					1	\Box						J		_]	_]		⅃			_]	J	\Box
G. astutoides STRAND 1946							\Box				1		_]	_]			J				J	

locality	Ti	2	3	4	5	6	7	R	,	10	111	12	13	14	15	16	17	18	19	20	21	22
G. exspectatus SMETANA 1951	Ť	Ī	Ť	Ė	Ť	Ė	Ė	Ť	ĺ	Ť	Ë			2		-				Ħ	1	=
Gabrius sp. 0	t	1	T	T	H	Г	Г	H	\vdash	T	H	H		۲	H	┢	H	H			H	
Ocypus picipennis (F. 1792)	1	Ė	Г	T	T	Г	Г	Г	T	T	Г	T	ī	Г	T	Г	T	П			П	H
Heterothops minutus WOLL.1860	\dagger	Г	Г		┢	Т			Т				Ė	Г	Τ	Г		6		П	П	П
Quedius microps GRAV. 1847	1	T	Г	1		Г		Г	Г	Г	Г	Г		Г	T	┢		Ť		П		
Q. mesomelinus (MARSH. 1802)	\top	Г	Г	Ė	Г	Г		Г	Г	П	Г	Г	Г	Г	Г	Г	ı	П		П		П
Q. xanthopus ER. 1839	\top	Г	Г		Г	Г	Г		Г	Г	Г	Г		Г						1		П
Q. lateralis (GRAV. 1802)	1	Г						1	Г	Г	П	П		Г	Г		Г					П
Q. bernhaueri RAMBOUSEK 1915	\top	Г	Г	Г	Т	Г		T	Г	Г				Г	Г		3					
Q. umbrinus ER. 1839	1		Г	Г	Г					13					Г				2	9	2	
Q. suturalis Kiesenw. 1847	1		Г	Г	Г		П	Г			Г	Г		Г	Г	Г	Τ				П	3
Q. erinci KORGE 1971	\top	Г	Г	Г	Г	Г	П	Г	Г	Г	Г			5	Г	2	Г			1		Ť
Q. pseudonigriceps REITT. 1909	T	ı	T		Г	Г	13			T	Г	Г		Ė		Ť	40			3	12	2
Q. pseudopyrenaeus Coiff. 1967	T	Ė	Г	Г		Г		Г	Γ	П	П			П		Г	H			6	Ħ	Ī
Q. cf. paganettii Bernh. 1936	╁	1	Г			Г	П	Г	_		Г	Г		Г	Г		П			Ť		П
Q. cinctus (PAYKULL 1790)	十	Ė	Г		_	Г	П	Г	Г		Т	П		П	Г		П				5	
Q. boops (GRAVENHORST 1802)	T	Г	Г		_						Н	П				4	4		3		۲	
Quedius spp. 00	1	1	Г	Г	_			1	Г		1	П		1	П		28				1	\neg
Habrocerus pisidicus KORGE 1971	T	Ė	Г	Г				4		Г		П			П							5
H. capillaricornis (GRAV. 1806)	T	Г	Г				Г	Г	3	35	П	П				Γ.	П					2
Mycetoporus bosnicus LUZE 1901	T	Г	Г	Г			П	1				П					П	П	1	П		٦
M. baudueri MULS.& REY 1875	1		Г	П			П		П	П				П	П	$\overline{}$	1					╗
M. cf. erichsonanus FAGEL 1965	T		2	1			П			П		П					П					П
M. cf. bosnicus OO	T		Г	П						П		1	1			2	1					1
M. forticornis FAUVEL 1872	Т		Г									П	7			_	П					
M. cf. nigricollis STEPH. 1835 оо	1									П				1				П		T		
Mycetoporus brucki (PAND. 1869)	Т						6										1			T	٦	
Ischnosoma longicorne (MAKLIN 1847)	Т	1							2								3	T		1	1	
Lordithon trinotatus (ER. 1839)		1				1	2	2			1	3		8			1					
L. thoracicus (F. 1777)																		\neg		1	\Box	
L. lunulatus (L. 1761)																	1			10	6	
Sepedophilus testaceus (F. 1792)								1						5			1					
S. immaculatus STEPH. 1832									1											1		
S. binotatus (GRAV. 1802)	Π																	\Box		1		
S. cf. apfelbecki (LUZE 1902)	1		1										7									
Tachyporus nitidulus (F. 1781)							4				1	2										ı
Tachinus corticinus GRAV. 1802																		1				
T. laticollis GRAV. 1802																		\Box		1		
T. bonvouloiri PAND. 1869	1										1		floor								\Box	
T. discoideus Er. 1839											3									floor		
Cypha sp.				\square								1				$_]$	1	J				
Holobus flavicornis (BOIS. & LAC. 1835)	L		Ш	Ш		Ш					\Box		1						╝		\perp	╝
Gyrophaena sp. Q	L	Ш								1	╝					$_{\perp}$						╝

locality	1	2	3	4	5	6	7	8	9	10	111	12	13	14	15	16	17	18	19	20	21	22
Deinopsis erosa (STEPH. 1832)	Ī	Г	Т			Т	Γ	Τ	Г	1	Г	Г		Г	Г			Π	Г	Г		Γ
Myllaena kraatzi SHARP 1871	T	Г	П		7		Г	Τ		Ť	Г	Г							Г	Г	Г	Г
M. aff. minuta (GRAV. 1806)	Π	1	П	П			Γ	Г	Г	10	Г					П	Г	Г	Г	П	П	П
Myllaena sp.							Г	Г	Г	ı		Г							Г			Г
Leptusa asperata EPPELSH. 1888	Г			2			Г	3	Г					20					Г		8	П
L. ruficollis (ERICHSON 1839)	Γ						Г	ı	Г								Г	Г	Г			
L. jelineki PACE 1983	Γ					Г	Γ	Г	Г								136	Γ	Γ			
L. peristerica PACE 1989																					48	
L. winneguthiana PACE 1983	Γ							Γ									Г		Г		14	
L. sororella PACE 1989																						47
Rhopalocerina clavigera (SCRIBA 1859)							1			1												
Bolitochara obliqua ER. 1837																			_	1	7	
Autalia rivularis (GRAV. 1802)																						1
Cordalia obscura (GRAV. 1802)			1																			
Anaulacaspis laevigata (EPPELSHEIM 1883)								Г		1												
Tachyusa balteata (ER. 1839)	Г				1			Γ		П							Г					
T. cf. coarctata (ER. 1837)					1																	
T. objecta (MULS. & REY 1870)	Γ				1																	
T. constricta (Er. 1837)	Г				1			Г														П
Ischnopoda umbratica (ER. 1837)	Г				1			Г														
Hydrosmecta sp.	Г						Г		П						21							
Aloconota mediterranea (BENICK 1941)																						1
Geostiba oertzeni (EPPH. 1888)							91				13	45										
G. obtusangula ASSING 2000		2						5														
G. torisuturalis Assing 2000																35		29				
G. fthiotisensis ASSING 1999	50												33									
G. breviuter ASSING 2000																				9		
Paraleptusa wunderlei Assıng 2000	4																					
Amischa strupii SCHEERP. 1967																			136	2		
A. bifoveolata (MANNERH.1830)																	1					\Box
Lyprocorrhe anceps (Er. 1837)																	16				\Box	
Notothecta flavipes (GRAV. 1806)															\exists		14			\Box		
Liogluta longiuscula (GRAV. 1802)					11	22	228	2		1	19	28	7				1		13	4		1
L. alpestris (HEER 1939)																	1			2		\exists
Emmeostiba? sp.							1															
Atheta elongatula (GRAV. 1802)					5																	\Box
A. brisouti (HAROLD 1867)																			4			
A. debilis (ER. 1837)																			1			
A. (Bessobia) sp.										\Box							1		2			
4. benickiella Brundin 1948		1			\Box		12			J		_]	_]		_]	_]	22	_]	_7	_]	2	_]
A. pittionii SCHEERPELTZ 1950										\Box			\Box						╛	J	2	\Box
A. (Microdota) sp.			\Box		\Box						\Box	_]				\Box		2	╝			\Box
A. sodalis (ER. 1837)		\Box												\Box					J	3		\Box

locality	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
A. gagatina (BAUDI 1848)	Γ			Г			Г	Γ	Г	Γ	Ī						Γ		Γ	Γ	Γ	2
A. bosnica GANGLBAUER 1895	T			Г	П			Γ		Γ	Γ								19			П
A. (Mocyta) spp.	Γ	1		13	Г		Г	3	Γ	Г	Г					1	ı			1	Г	П
A. hypnorum (KIESENW. 1850)	Γ			Г				Г	Г	Г		П								1	Γ	П
A. crassicornis (F. 1792)				4	Г			Г	Γ	Г		Г	ı							Г	1	П
A. marcida (ER. 1837)								Г	Г	Г		Г					Г			Г	1	П
A. putrida (KRAATZ 1856)	Γ	6				3	32		Г	Г	1	ı					2			22	1	П
Atheta sp.												П	1									П
Acrotona muscorum (BRIS.1860)	Г										1											П
A. consanguinea (EPPH. 1857)	Г	3												5								П
A. parens (MULS. & REY 1852)	Г							Г							П			2			2	П
A. nigerrima (AUBÉ 1850)	Г						Г								П	Г	П		1			П
Zyras collaris (PAYK. 1800)										1												П
Cousya cf. nitidiventris (FAGEL 1958)	Г			П							Г										1	П
Cousya sp.	Г		1								Г			1								П
Meotica sp.		1											2									П
Amarochara forticornis (Bois. & Lac. 1835)						1																
Parocyusa longitarsis (ER. 1837)					124					3	1		6		1							
Tectusa viduus sp. n.	2															П		\neg				П
T. timfristosensis sp. n.	6																	П				
T. longiuter sp. n.				2																		
T. apollonis ZERCHE in prep.							7				10											
T. rastrifera sp. n.							4					40										
T. vardousiensis sp. n.													11									
T. sp. n.																3						
Ocalea robusta BERNH. 1902									1													1
O. concolor Kiesenw. 1847										1												
O. puncticeps KRAATZ 1858																	9			12	12	
O. cf. badia ER. 1837								1												1		
Ischnoglossa prolixa (GRAV. 1802)								1														
Oxypoda opaca (GRAV. 1802)		╝			╝	_	╝					┙	1					╝				╝
O. ignorata ZERCHE 1996			_	╝	╝	\perp		╝	_	╝	Ц						1	╝	╝		1	
O. attenuata MULS. & REY 1851				\Box		\Box		1	_	╝							╛		\Box			
O. vicina Kraatz 1856			2																			
O. brevicornis (STEPH. 1832)			_		\perp			_	╛	╝				\perp		2			1	2	$_$ $[$	1
O. mutata Sharp 1871				1	┙	Ц		╝	_	┙	\sqcup			\perp		_			_			╛
O. mulsanti Bernhauer & Scheerpeltz 1926	\Box	\perp	_[\bot	\perp	\perp	\perp	_[\perp	_[\perp	\perp	\perp	ij	\Box	\Box		\perp		\Box	\bot	_]
O. aff. brachyptera (Sтерн. 1832)	2	\perp		\Box	_	_[\perp	_	\Box	_	1	⅃	1	\perp	\perp	ot	_[$oldsymbol{ol}}}}}}}}}}}}}}}}}}}}}$		$ \bot $	\prod	╝
O. annularis MANNERH. 1830		ot	⅃	\Box	ightharpoonup	ot	\perp	\perp	ot	\Box		⅃	\perp	\perp	\perp		69	\rfloor		20	14	
O. flavicornis KRAATZ 1856		_	_	_	_	_	Ц	_	_	╝		┙	\perp	\perp	\perp	\perp	3	\perp		Ц	_[
O. lurida WOLLASTON 1857		ightharpoonup	_		_	Ц	Ц	1	2	2			⅃	\perp	\perp	\prod	\perp	⅃	╝	\perp	_[\Box
O. recondita KRAATZ 1856		_	ļ		_	1	1	_	_	ļ	_	1	\Box	⅃	\perp	\perp	9	\perp		5	_[╝
O. (Baeoglena) sp.				\perp	\Box	\perp	\perp					1		1	_[_[12			6	6	╝

locality	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
Oxypoda sp. 1	T.						1															
Oxypoda sp. 2		L	Ŀ					1														
Oxypoda sp. 3		L												1								
Homoeusa acuminata (MÄRKEL 1842)								1														
Haploglossa villosula (STEPHENS 1832)							1															
Crataraea suturalis (MANN. 1830)																	1					
Aleochara aff. laevigata GYLLENHAL 1810	5																					
A. erythroptera GRAV. 1806	\mathbb{T}_{-}						1															
Aleochara sp.											2								14			

Remarkable records and descriptions of new species

Anthobium ganglbaueri (LUZE 1905)

T y p e s e x a m i n e d : Syntypes: 1δ : δ / Süd-Ungarm / Ganglbaueri m. Luze / ex coll. Luze / TYPUS Lathrimaeum Ganglbaueri Luze; 1δ : δ / Ganglb. 95 Herkulesbad / coll. Schuster / COTYPUS Lathrimaeum Ganglbaueri Luze (Naturhistorisches Museum Wien).

A d d i t i o n a 1 m a t e r i a 1 e x a m i n e d: 13, Romania, Herkulesbad, leg. Winkler (Naturhistorisches Museum Wien); 13, Greece, Makedhonia, Pieria Ori, Katafygi, 40°16′20N, 22°09′00E, 1450m, *Pinus* forest, 9.IV.1998, leg. Assing (cAss).

A. ganglbaueri, originally described from Romania and later also reported from Bulgaria and Yugoslavia (FAGEL 1968, SCHEERPELTZ 1961) is here for the first time recorded from two localities in Greece (Pieria, Florina), where it was sifted from litter in a Pinus forest and in an oakwood at altitudes of 1450 and 850m, respectively (Tab. 1 and additional material examined).

Deliphrosoma angulatum sp. n. (Figs. 1-3)

Holotype ♂: GR. Fokis, No. 17, 45km SW Lamía, Oros Vardousia, 1600m, 38°42′06N, 22°08′54E, 18.IV.2000, V. Assing (cAss).

P a r a t y p e s: 10: same data as holotype (cAss); 1033, 10: Zentral-GR, Parnassosgebirge, 2250 Gesiebe Gräser, Schneefeldrand, 11.6.1996 P. Wunderle (cAss, cWun); 13: GR. Fthiotis, No. 9, Parnassos Oros, 1700m, Abies wood with snow, 38°34′12N, 22°34′24E, 15.IV.2000, V. Assing (cAss); 200; GR. Fokis, No. 15, 45km SSW Lamía, Oros Giona, 1700m, 38°39′36N, 22°19′32E, 17.IV.2000, V. Assing (cAss); 13, 200; same data, but leg. Wunderle (cWun).

Description: 4.2-5.0 mm. Head, pronotum, and abdomen blackish brown to black, with the narrow lateral margins and the elytra somewhat ligher, dark reddish brown; legs bicoloured, with the femora dark brown, and the tibiae and tarsi yellowish brown; antennae dark brown to blackish brown, with antennomere I usually somewhat lighter, reddish brown to brown; palpi brown to blackish brown.

Head as in *D. fratellum* (ROTTENBERG), but usually with a shallow furrow between ocelli. Pronotum similar to that in *D. fratellum*, but with more narrowly and less distinctly separated lateral margins. Elytra and abdomen as in *D. fratellum*.

3: protibia in middle with distinct angular dilatation (Fig. 2); aedeagus of similar shape as in D. fratellum, but tube in internal sac with more coils, proximally less strongly dilated, and more distinctly sclerotized in the middle (Figs. 1, 3).

Derivation of minis: The name (adj.) refers to the angular dilatation of the δ protibia, which readily distinguishes the new species from D. fratellum.

Comparative notes: From D. fratellum, according to ZERCHE (1991) the only known Greek congener, D. angulatum is separated especially by the more distinctly modified & protibiae (Figs. 2, 4), the internal structures of the aedeagus (Figs. 3, 5), the presence of a shallow furrow between the ocelli, the usually lighter antennomere I, and the narrower and less distinctly separated lateral margins of the pronotum. It should be pointed out that D. fratellum is usually of darker colour than indicated in the redescription by ZERCHE (1991); this particularly applies to the pronotal margins and antennomere I. (68 specimens of D. fratellum from Greece and Yugoslavia were examined.) In the similar D. pirinense ZERCHE 1991 from Bulgaria, the & protarsi are weakly modified and the morphology of the aedeagus is different. For distinguishing characters and illustrations of the aedeagi of other species occurring in the Balkans, the descriptions, key, and drawings in ZERCHE (1991, 1998) are referred to.

Distribution and bionomics: D. angulatum is known from three mountain ranges (Vardousia, Giona, Parnassos) in southern central Greece (Fokis, Fthiotis), where it was sifted from grass, litter of shrubs, and from litter in an Abies forest, mostly near snow, at altitudes between 1600 and 2250m.

Pareudectus vitsiensis sp. n. (Figs. 6-10)

Holotype &: GR. Florina, No. 20, 15km S Flórina, Oros Vitsi, 1900m, 40°39N, 21°23E, 21.IV.2000, V. Assing (cAss).

P a r a t y p e s: 1 o same data as holotype, but leg. Wunderle (cWun); 2 o o same data, but 25.IV.2000, leg. Assing, Wunderle (cAss, cWun).

Description: Measurements (in mm) and ratios (range; n=4): HL: 0.41-0.45; HW: 0.50-0.54; PL: 0.47-0.54; PW: 0.54-0.60; EL: 0.39-0.45; ML: 0.60-0.65; TL: 2.8-3.6; HL/HW: 0.82-0.85; HW/PW: 0.89-0.92; PL/PW: 0.86-0.89; EL/PL: 0.80-0.85.

Coloration uniformly ferrugineous. Facies as in Fig. 9. Head of similar outline as in *P. apfelbecki* (GANGLBAUER); eyes slightly shorter than postgenae in dorsal view; puncturation coarse and relatively dense, in lateral dorsal area (near eyes) rugosely confluent; integument without distinct microsculpture. Antenna as in Fig. 10.

Pronotum wider than head and somewhat transverse (see ratios HW/PW and PL/PW), with crenulate, narrowly separated lateral margins; puncturation relatively coarse, interstices shining, without microsculpture; in anterior half with shallow ovoid median impression; in posterior half of midline with impunctate, shining longitudinal elevation.

Elytra at suture shorter than pronotum (see ratio EL/PL), but distinctly exceeding the latter in width (see measurements and Fig. 9), anterior angles more or less rounded, but distinct (similar to *P. leonhardi* (BERNHAUER)); posterior angles obliquely truncate; puncturation coarse, somewhat denser and more irregularly spaced than that of pronotum; interstices with very shallow microsculpture.

Abdomen very wide, much wider than elytra (see Fig. 9), distinctly dilated posteriorly, maximal width across segments V/VI; puncturation much finer and sparser than that of forebody, weakly granulose; integument mat due to distinct microreticulation.

3: outline of sternum IX as in Fig. 8; aedeagus with ventral process long, slender, and apically rounded; internal sac with two long rows of numerous distinctly sclerotized spines (Figs. 6-7).

D e r i v a t i o n o m i n i s: The name is derived from the type locality of the species, the Oros Vitsi in northern Greece.

C o m p a r a t i v e n o t e s: Only three species of Pareudectus ZERCHE 1990 were previously known: P. eppelsheimi (GANGLBAUER 1896) from the Carpathians, P. apfelbecki (GANGLBAUER 1892) from Bosnia-Herzegovina, and P. leonhardi (BERNHAUER 1905) from Bosnia-Herzegovina and Montenegro. P. vitsiensis sp. n. is distinguished from all known congeners by the shape and internal structures of the aedeagus (for comparison see figures in ZERCHE (1990)), from P. eppelsheimi and P. leonhardi in addition by larger size and larger eyes, and from P. apfelbecki, in which the internal structures of the aedeagus are somewhat similar, by the more transverse pronotum, more distinct anterior angles of the elytra, and by the narrower abdomen.

D is tribution and bionomics: The new species is the first representative of the genus from Greece. It was collected on the northwestern slope of the peak of the Oros Vitsi (Florina), where it was sifted from grass roots and mosses near big rocks at an altitude of 1900m, together with a new species of *Coryphiodes* BERNHAUER (ZERCHE in prep.), and one of *Geostiba* THOMSON (ASSING 2000).

Lathrobium (s. str.) vorasensis sp. n. (Figs. 11-16)

Holotype &: GR. Florina, No. 26, 40km NE Flórina, Oros Voras, 1650m, 40°52′58N, 21°47′36E, 22.IV.2000, P. Wunderle (cWun).

Paratype o: same data as holotype, but leg. Assing (cAss).

Description: Measurements (in mm) and ratios (holotype, paratype): HL: 0.80, 0.73; HW: 0.73, 0.66; PL: 0.97, 0.87; PW: 0.73, 0.64; EL: 0.59, 0.57; ML: 1.25; TL (from mandibles to hind margin of tergum VIII): 5.7, 5.5; HL/HW: 1.09, 1.10; HW/PW: 1.01, 1.02; PL/PW: 1.33, 1.35; EL/PL: 0.61, 0.66.

In external appearance (proportions, coloration, puncturation) similar to the only known microphthalmous Greek congener *L. wunderlei* ASSING & SCHÜLKE 2000, but body size larger (see measurements). Microsculpture of head distinctly isodiametric. Eyes as reduced as in *L. wunderlei*, rudiments without distinct ommatidia. Head only slightly wider than pronotum (see ratio HW/PW).

- δ: sternum VII posteriorly with condensed dark and long pubescence, hind margin broadly concave; hind margin of tergum VIII unmodified, weakly convex; sternum VIII posteriorly with pronounced U-shaped incision, in postero-median area with dense long and dark setae directed diagonally medio-caudad, lateral parts of hind margin on either side of incision distinctly projecting caudad (Fig. 16); aedeagus as in Figs. 11-13.
- q: posterior margin of tergum VIII weakly convex, sternum VIII posteriorly with blade-like projection (Fig. 15); dorsal aspect of segments IX and X as in Fig. 14.

Derivatio nominis: The name refers to the type locality of the species, the Voras Oros in northern Greece.

C o m p a r a t i v e n o t e s : From L. wunderlei, L. vorasensis is readily separated by its larger size, the relatively wider pronotum, the different shape and chaetotaxy of the δ sternum VIII, the shape and internal structures of the aedeagus, the absence of an acute projection at the hind margin of the φ tergum VIII, and by the different shape of the φ sternum VIII; for illustrations and a description of L. wunderlei see ASSING & SCHÜLKE 2000). The morphology of the aedeagus (external shape, internal structures) is most

similar to that in *L. matchai* RAMBOUSEK from the nearby Pelister in Macedonia. In that species, however, the ventral process of the aedeagus is apically acute (and not truncate) and more slender in ventral view, and the dorsal sclerite is apically shorter and less acute. For figures of the aedeagi of *L. matchai* and other micro- and anophthalmous species of *Lathrobium* s. str. occurring in the Balkans see PACE (1984).

Distribution and bionomics: The new species is known only from the Voras Oros in northern Greece, very close to the Macedonian border, where it was sifted from moist leaf litter in a beechwood in northern exposition at an altitude of 1650m.

Sunius fokisensis sp. n. (Figs. 17-18)

H o I o t y p e δ : GR. Fokis, No. 17, 45km SW Lamia, Oros Vardousia, 1600m, 38°42′06N, 22°08′54E, 18.IV.2000, V. Assing (cAss).

Paratypes: 950, 1200: same data as holotype (cAss); 10, 400: same data as holotype, but leg. Wunderle (cWun).

Description: Very small species, 2.7-3.2 mm (abdomen extended); head width 0.37-0.42 mm. Forebody rufous, with the head and the elytra sometimes slightly infuscate; abdomen, except for the lighter apex, dark brown to blackish; antennae, legs, and palpi testaceous.

Head approximately as long as wide; postgenae in dorsal view subparallel, nearly three times as long as eyes; puncturation coarse, but sparse; interstices shining, without microsculpture.

Pronotum slightly narrower than head and slightly longer than wide; puncturation as coarse as that of head, but somewhat denser, midline broadly impunctate; interstices shining, without microsculpture.

Elytra subequal in width to pronotum, but distinctly shorter, at suture ca. 0.7x the length of pronotum; puncturation rather dense, serrate, and weakly granulose; interstices without microsculpture; hind wings reduced.

Abdomen with very fine puncturation and shallow microreticulation composed of relatively wide meshes.

 δ : sternum VIII triangularly incised posteriorly; aedeagus distinctive, slender, and with characteristic sclerotized structure in internal sac (Figs. 17-18).

Derivatio nominis: The name is derived from Fokis, the province in which the Vardousia Oros, the type locality, is located.

C o m p a r a t i v e n o t e s: Four species of Sunius CURTIS 1829 have been reported from Greece or can be expected to occur there: S. melanocephalus (FABRICIUS 1792), S. fallax (LOKAY 1919), S. hellenicus (COIFFAIT 1961), and S. bicolor (OLIVIER 1795). All these congeners are distinguished from S. fokisensis by larger size (head width: >0.44 mm, usually distinctly larger) and by the completely different morphology of the aedeagus. For illustrations see COIFFAIT (1984), but note that S. fallax sensu COIFFAIT (1984) refers to a different species; the aedeagus of S. fallax (LOKAY) is illustrated by ASSING (1995).

Distribution and bionomics: S. fokisensis is known only from the Vardousia Oros, Fokis, where most specimens were sifted from grass and grass roots, some also found under stones, together with Geostiba fthiotisensis ASSING, Tectusa

vardousiensis sp. n., and an anophthalmous species of Caecoparvus JEANNEL (Carabidae, Bembidiinae) at an altitude of 1600m.

Quedius bernhaueri RAMBOUSEK 1915 (Figs. 19-22)

This remarkable endogean species was described from the Pelister range in Macedonia, but has apparently not been recorded again. RAMBOUSEK (1915) originally attributed it to the new subgenus *Tenebrobius*, but today it is referred to the subgenus *Microsaurus* DEJEAN (COIFFAIT 1978).

We sifted three specimens of the species near the peak of the Oros Vitsi, Florina, from deep leaf litter in a beechwood on calcareous soil in northern and northwestern exposition at altitudes of 1800 and 1900m, together with a possibly new anophthalmous species of Lathrobium (Tab. 1). Q. bernhaueri is here for the first time recorded from Greece.

Leptusa jelineki PACE 1983

The species was previously known only from the type locality, the Galičica planina in Macedonia (PACE 1989) and his here for the first time reported from Greece. We collected the species in large numbers near the peak of the Oros Vitsi, Florina, in a calcareous beechwood in northern and northwestern exposition at an altitude of 1800-1900m (Tab. 1). In addition, there is a record of one \eth from the Vermio Oros (Imathia), NW Kato Vermio, 1300m, 6.IV.1999, leg. Brachat (cAss).

Leptusa peristerica PACE 1989

Like the preceding species, L. peristerica, which was known only from the type locality, the Pelister range in Macedonia, is evidently more widespread. We sifted it in the Varnous Oros, ca. 15 km W Florina, from leaf litter in a beechwood at an altitude of 1600m (Tab. 1), together with the following species. This is the first record from Greece.

Leptusa winneguthiana PACE 1983

Types examined: <u>Holotype</u> δ : δ / Golešnica-pl. Wgth. 28.7. / ex coll. Scheerpeltz / TYPUS Leptusa Winneguthiana O. Scheerpeltz / HOLOTYPUS Leptusa (Stictopisalia) winneguthiana m. det. R. PACE 1980 (NHMW). <u>Paratype</u> δ : same labels as holotype, but "PARATYPUS ..." (Naturhistorisches Museum Wien).

According to PACE (1989), L. winneguthiana is endemic in the Golešnica planina in southern Serbia. The present first record from Greece, however, shows that the species is much more widely distributed. We found 14 specimens (compared with the types) in the same locality and under the same circumstances as the preceding species in the Varnous Oros, Florina (Tab. 1).

Leptusa sororella PACE 1989

Like the three preceding species, L. sororella was previously known only from the type locality, Prespa Jezero in Serbia (PACE 1989). The present first record from Greece shows that its range of distribution is obviously much more extensive. Numerous specimens were sifted from leaf litter in an oakwood, ca. 10 km south of Flórina, at an altitude of 850m (Tab. 1).

Tectusa rastrifera sp. n. (Figs. 23-32)

Holotype ♂: GR. Fokis, No. 16, 45km SSW Lamia, Oros Giona, 1650m, 38°39'36N, 22°19'12E, 17.IV.2000, V. Assing (cAss).

Paratypes: 433, 1000: same data as holotype (cAss); 933, 1600: same data, but leg. Wunderle (cWun); 233, 250: GR. Fthiotis, No. 9, Parnassos Oros, 1700m, Abies wood with snow, 38°34′12N, 22°34′24E, 15.IV.2000, V. Assing (cAss).

D e s c r i p t i o n : Measurements (in mm) and ratios (range, arithmetic mean; n=19): AL: 0.89-1.03, 0.95; HW: 0.41-0.46, 0.43; PL: 0.42-0.49, 0.45; PW: 0.48-0.57, 0.53; EL: 0.30-0.35, 0.33; HTi: 0.44-0.53, 0.48; HTa: 0.30-0.38, 0.34; ML: 0.48-0.51, 0.50; TL: 2.7-3.6, 3.2; PW/HW: 1.17-1.28, 1.22; PW/PL: 1.12-1.23, 1.16; EL/PL: 0.68-0.74, 0.72; HTa/HTi: 0.68-0.77, 0.72.

Rather large species. Coloration variable, bicoloured, with the forebody reddish brown and the abdomen (except for the lighter apex and tergal margins) blackish brown to almost uniformly blackish brown; antennae rufous; legs rufous to castaneous.

Head with lateral outline smoothly convex; postgenae in dorsal view nearly three times as long as eyes; eyes weakly projecting from lateral outline; puncturation very fine, barely noticeable in the distinct transverse microsculpture; pubescence relatively long and suberect. Antennae relatively long and slender, reaching the anterior third of elytra; antennomeres I-III distinctly oblong, III somewhat shorter than II, IV and V subquadrate, VI-IX increasingly, but relatively weakly transverse, X almost 1.5x as wide as long, and XI shorter than the combined length of IX and X (Fig. 32).

Pronotum distinctly wider than head and wider than long (see ratios PW/HW and PW/PL), more strongly narrowed cephalad than caudad, and distinctly convex in cross-section; puncturation more pronounced than that of head, relatively dense and finely granulose; integument with microreticulation and weak shine; pubescence similar to that of head.

Elytra clearly shorter (see ratio EL/PL) and slightly wider than pronotum; hind margin distinctly sinuate near posterior angles; puncturation more pronounced and more distinctly granulose than that of pronotum; microsculpture predominantly composed of diagonal long meshes; pubescence shorter than that of pronotum, directed caudad. Hind wings reduced. Legs relatively long, metatarsi not much shorter than metatibiae (see ratio HTa/HTi), first metatarsomere of somewhat variable length, but in most specimens approximately as long as the combined length of the three following tarsomeres or nearly so.

Abdomen subparallel, weakly dilated caudad, maximal width across segment V; terga III-V with pronounced, tergum VI with weaker anterior impression; puncturation fine and moderately dense; microsculpture composed of short transverse, almost isodiametric meshes.

- 3: tergum VIII posteriorly weakly pointed in the middle (Fig. 28); posterior margin of sternum VIII strongly convex (Fig. 29); aedeagus large, median lobe with distinctive structures in internal sac (Figs. 23-25); apical lobe of paramere as in Fig. 26.
- Q: tergum VIII similar to that in ♂ (Fig. 30); posterior margin of sternum VIII more weakly convex, with row of distinctly modified setae, and with very short, sparse, and rather stout micropubescence (Fig. 31); spermatheca as in Fig. 27.

Derivation ominis: The name refers to the distinctive apical internal structure of the aedeagus, which somewhat resembles a hoe or a mattock (Lat.: rastrum).

C o m p a r a t i v e n o t e s a n d c o m m e n t s: A comparison with similar congeners is somewhat problematic, since numerous species, particularly from the southern Balkans where the genus is highly diverse, have not yet been described or transferred to *Tectusa* BERNHAUER 1899. A comprehensive revision is currently in progress (ZERCHE in prep.). The only species previously attributed to *Tectusa* is the type species, *T. difficilis* (EPPELSHEIM 1880) from Bosnia, whose types were examined by the second author. *T. difficilis* is externally similar to the new species, but distinguished by larger size, smaller eyes, the weakly impressed pronotal midline, the more distinctly dilated abdomen, and the different morphology of the sexual characters.

It is worth noting that the specimens from the Parnassos are somewhat darker than those from the Giona Oros; however, no differences were found in the primary and secondary sexual characters.

D is tribution and bionomics: In contrast to many other Greek congeners, the distribution of *T. rastrifera* is not confined to a single mountain range. We sifted a total of 44 specimens both in the Parnassos Oros (Fthiotis) and in the Giona Oros (Fokis) from litter of *Abies* forests, partly near snow, at altitudes of 1700 and 1650m, respectively.

Tectusa longiuter sp. n. (Figs. 33-36)

Holotypeo: GR. Evritania, Nr. 4, 13 km SSW Karpenisi, Mt. Kaliakouda, 1200m, 38°49′00N, 21°46′40E, 14.IV.2000, P. Wunderle (cWun). Paratype: 10: same data as holotype, but leg. Assing (cAss).

Description: Measurements (in mm) and ratios (range; n=2): AL: 0.83-0.86; HW: 0.41-0.42; PL: 0.43-0.44; PW: 0.50-0.51; EL: 0.27-0.28; HTi: 0.40-0.42; HTa: 0.29-0.32; TL: 3.0-3.4; PW/HW: 1.22-1.24; PW/PL: 1.16-1.17; EL/PL: 0.63-0.64; HTa/HTi: 0.69-0.81.

Of similar size (see measurements) and external appearance (proportions, coloration, puncturation, microsculpture) as *T. rastrifera*, but distinguished as follows:

Head with smaller eyes, the latter not projecting from lateral outline of head, postgenae in dorsal view at least three times as long as diameter of eyes. Antennae shorter (see measurements) and with more transverse antennomeres X (Fig. 33).

Elytra shorter than in *T. rastrifera* (see measurements and ratios); metatarsi shorter, metatarsomere I at most only slightly longer than the combined length of the two following metatarsomeres.

♂: unknown.

q: tergum and sternum VIII similar to those in *T. rastrifera* (Figs. 35-36); spermatheca with distinctly longer and more slender duct, and with relatively smaller capsule (Fig. 34).

Derivatio nominis: The name is a noun in apposition composed of the Latin adjective longus and the noun uter (hose, tube) and refers to the longe duct of the spermatheca, which distinguishes this species from the similar *T. rastrifera*.

C o m p a r a t i v e n o t e s: For separation from *T. rastrifera* sp. n. and *T. difficilis* see the description above and the comparative notes below *T. rastrifera*. The following species are readily distinguished by much smaller body size alone.

Distribution and bionomics: The types were collected in the

Kaliakouda range, Evritania, by sifting litter in a firwood at an altitude of 1200m.

Tectusa viduus sp. n. (Figs. 37-42)

Holotype &: GR. Evritania, W Lamia, N Karpenisi, Timfristos, ski resort, 1900m, 38°57N, 21°48E, No. 1, 14.IV.2000, P. Wunderle (cWun).

Paratype: 18: same data, but leg. Assing (cAss).

Description: Measurements (in mm) and ratios (range, n=2): AL: 0.74-0.76; HW: 0.38; PL: 0.34-0.35; PW: 0.38-0.41; EL: 0.21-0.22; HTi: 0.35; HTa: 0.21-0.22; ML: 0.35-0.36; TL: 2.6-2.9; PW/HW: 1.00-1.08; PW/PL: 1.11-1.17; EL/PL: 0.61-0.64; HTa/HTi: 0.61-0.63.

Forebody reddish brown; abdomen blackish brown, with the tergal margins and the apex lighter; antennae rufous, legs and palpi yellowish brown.

Head subcircular with distinctly convex lateral outline; eyes small, not projecting from lateral outline of head, postgenae in dorsal view more than four times as long as eyes; puncturation very fine; integument with microreticulation and almost mat. Antennae with antennomeres I and II oblong and of equal length, III subquadrate, IV-IX increasingly transverse, X almost twice as wide as long, and XI rather short, distinctly shorter than the combined length of IX and X.

Pronotum weakly transverse, not or only weakly wider than head; lateral margins in dorsal view convex; puncturation finely granulose, less fine and more distinct than that of head; microreticulation similar to that of head, but with finer meshes; pubescence decumbent, in anterior half of midline directed cephalad, in posterior half of midline caudad, and in lateral area transversely laterad.

Elytra distinctly shorter (see ratio EL/PL) and slightly wider than pronotum, not dilated posteriorly; anterior angles distinct; posterior margin obliquely truncate, near posterior angles with moderate concavity; puncturation slightly coarser than that of pronotum, but less dense; microsculpture similar to that of pronotum, but shallower, integument with subdued shine; pubescence shorter than that of pronotum, directed diagonally laterocaudad. Hind wings reduced. Tarsi relatively short; first metatarsomere shorter than the combined length of the two following tarsomeres; terminal metatarsomere approximately as long as the combined length of the three preceding tarsomeres.

Abdomen weakly dilated caudad, maximal width across segments VI/VII; terga III-V with distinct, tergum VI with weaker anterior impression; puncturation very fine, barely noticeable in the microsculpture, which is composed of short transverse meshes.

 δ : tergum VIII posteriorly weakly convex (Fig. 41); posterior margin of sternum VIII in the middle obtusely pointed and with long setae (Fig. 42); aedeagus with ventral process of median lobe relatively long and strongly bent (Figs. 37-39); apical lobe of paramere as in Fig. 40.

g: unknown

Derivatio nominis: The name (Lat.: widower) is a noun in apposition and refers to the fact that the females of this species are unknown.

C o m p a r a t i v e n o t e s: T. viduus is readily separated from the two preceding species and from T. difficilis by external characters alone: the distinctly smaller body size, the more reduced eyes, the more pronounced microsculpture and the less dense pubescence. In addition, it is distinguished by the morphology of the primary and secon-

118

dary sexual characters.

D is tribution and bionomics: The types were collected near the ski resort of the Timfristos Oros, Evritania, on a slope in northern exposition, where they were sifted from grass roots, moss, etc. near snow at an altitude of 1900m, together with the following species.

Tectusa timfristosensis sp. n. (Figs. 43-51)

Holotype &: GR. Evritania, W Lamia, N Karpenisi, Timfristos, ski resort, 1900m, 38°57N, 21°48E, No. 1, 14.IV.2000, P. Wunderle (cWun).

P a r a t y p e s : 3 Q Q: same data as holotype (cWun); 2 Q Q same data, but leg. Assing (cAss).

Description: Measurements (in mm) and ratios (range; n=6): AL: 0.59-0.63; HW: 0.29-0.31; PL: 0.29-0.32; PW: 0.30-0.34; EL: 0.17-0.18; HTi: 0.26-0.29; HTa: 0.17-20; ML: 0.28; TL: 2.0-2.3; PW/HW: 1.05-1.13; PW/PL: 1.05-1.10; EL/PL: 0.55-0.60; HTa/HTi: 0.64-0.69.

Colour of body, except for the lighter abdominal apex, blackish brown, forebody sometimes lighter brown; legs and antennae yellowish brown.

Head subcircular with convex lateral outline, but less so than in *T. viduus*; eyes larger than in *T. viduus*, postgenae in dorsal view approximately three times as long as eyes; puncturation very fine, barely noticeable in the partly transverse and partly isodiametric microsculpture. Antennae with antennomeres I-III oblong, I and II of equal length, III shorter than II, IV-IX increasingly transverse, X ca. 2.5 times as wide as long, and XI short, distinctly shorter than the combined length of IX and X.

Pronotum weakly transverse, slightly wider than head; more distinctly tapering caudad and lateral margins less convex than in *T. viduus*; puncturation finely granulose, denser and more distinct than that of head; microreticulation composed of irregularly shaped meshes; pubescence similar to that in *T. viduus*.

Elytra much shorter (see ratio EL/PL) and indistinctly wider than pronotum, weakly dilated posteriorly; anterior angles indistinct; posterior margin truncate, near posterior angles weakly concave; puncturation moderately dense and rugose, coarser than that of pronotum; microsculpture with larger meshes than that of pronotum, integument with subdued shine; pubescence similar to that in *T. viduus*. Hind wings reduced. Tarsi relatively short; first metatarsomere shorter than the combined length of the two following tarsomeres; terminal metatarsomere shorter than the combined length of the three preceding tarsomeres.

Abdomen distinctly dilated caudad, maximal width across segments VI/VII; terga III-V with distinct, tergum VI with weaker anterior impression; puncturation very fine, but slightly more distinct than in *T. viduus* and weakly granulose; microsculpture composed of transverse meshes

δ: tergum VIII posteriorly weakly convex (Fig. 48); posterior margin of sternum VIII in the middle obtusely pointed and with moderately long setae (Fig. 49); aedeagus with ventral process of median lobe shorter and less strongly bent than in *T. viduus* (Figs. 43-45); apical lobe of paramere as in Fig. 46.

Q: tergum VIII similar to that in d (Fig. 50); sternum VIII indistinctly pointed posteriorly (Fig. 51); spermatheca as in Fig. 47.

Derivation ominis: The name is derived from the type locality of the species,

the Timfristos Oros in Evritania.

C o m p a r a t i v e n o t e s: T. timfristosensis is readily separated from the syntopic T. viduus by the smaller size and other characters indicated in the description above.

Distribution and bionomics: The species was collected under the same circumstances as T. viduus (see comments below that species).

Tectusa vardousiensis sp. n. (Figs. 52-60)

H o l o t y p e \eth : GR. Fokis, No. 17, 45km SW Lamía, Oros Vardousia, 1600m, 38°42′06N, 22°08′54E, 18.IV.2000, V. Assing (cAss).

Paratypes: 233, 400: same data as holotype (cAss); 13, 300: same data, but leg. Wunderle (cWun).

Description: Measurements (in mm) and ratios (range, n=11): AL: 0.60-0.71; HW: 0.31-0.36; PL: 0.27-0.36; PW: 0.33-0.42; EL: 0.18-0.22; HTi: 0.25-0.31; HTa: 0.16-0.20; ML: 0.32-0.35; TL: 2.3-2.9; PW/HW: 1.04-1.15; PW/PL: 1.08-1.22; EL/PL: 0.57-0.67; HTa/HTi: 0.60-0.68.

In general appearance similar to *T. viduus*, though on average smaller. Body bicoloured, with the forebody usually reddish brown or (more rarely) dark brown and the abdomen blackish, except for the lighter apex and tergal margins; antennae, legs, and palpi testaceous to rufous.

Head of similar outline as in *T. viduus*, but slightly narrower (see measurements); eye size as in *T. viduus*; puncturation fine and shallow; integument with microsculpture predominantly composed of short or isodiametric meshes, only with subdued shine. Antenna similar to that in *T. viduus*, but slightly shorter (see measurements) and antennomeres V-X slightly more transverse.

Pronotum weakly transverse and slightly wider than head (see ratios PW/HW and PW/PL); maximal width close to anterior angles; lateral margins in dorsal view less convex than in *T. viduus*, in posterior half almost straight; posterior angles usually more marked than in *T. viduus*; median line more or less shallowly impressed; puncturation and microsculpture similar to those of head; pubescence of similar pattern as in *T. viduus*, but somewhat longer.

Elytra shorter (see ratio EL/PL) and only indistinctly wider than pronotum, not dilated posteriorly; anterior angles marked, posterior margin obliquely truncate, near posterior angles with moderate concavity; puncturation finely granulose and slightly more distinct than that of pronotum, but less dense; microsculpture similar to that of pronotum, but with somewhat wider meshes; surface with subdued shine; pubescence directed diagonally latero-caudad. Hind wings reduced. Tarsi relatively short; first metatarsomere distinctly shorter than the combined length of the two following tarsomeres; terminal metatarsomere almost as long as the combined length of the three preceding tarsomeres.

Abdomen weakly dilated caudad, lateral margins of segments III-VII in dorsal view forming a remarkably straight line; maximal width across segments VI/VII; terga III-V with distinct, tergum VI with weaker anterior impression; puncturation fine; microsculpture predominantly composed of long transverse meshes.

3: tergum VIII posteriorly weakly convex (Fig. 57); posterior margin of sternum VIII in the middle obtusely pointed and with moderately long setae (Fig. 59); median lobe of aedeagus and apical lobe of paramere as in Figs. 52-55.

Q: tergum VIII similar to that in δ (Fig. 58); sternum VIII posteriorly convex (Fig. 60); spermatheca as in Fig. 56.

Derivation ominis: The name is derived from the type locality of the species, the Vardousia Oros in Fokis.

C o m p a r a t i v e n o t e s: From the externally similar, but somewhat larger T. viduus, T. vardousiensis is distinguished by the slightly narrower head, the shorter antennae, the shape of the pronotum, the shape and the microsculpture of the abdomen as well as by the shape of the aedeagus (see description of T. vardousiensis). The other species described or referred to above differ in the sexual characters and are either distinctly smaller and of darker colour (T. timfristosensis) or larger (T. rastrifera, T. longiuter, T. difficilis).

D is tribution and bionomics: The types were discovered in the Vardousia Oros, Fokis, on a slope in northern exposition, where they were sifted from litter of shrubs near snow and from grass at an altitude of 1600m, together with *Sunius fokisensis* sp. n., *Geostiba fihiotisensis* ASSING, and an anophthalmous species of *Caecoparvus* JEANNEL (Carabidae, Bembidiinae).

Tectusa vrontousensis sp. n. (Figs. 61-69)

H o l o t y p e δ : GR. (NE) Makedhonia, N Séres; Vrontous; 5, peak above Lailias, W-slope; 1800m, 25.V.1999, V. Assing (cAss). P a r a t y p e s: 1δ , 2_{Q} $_{Q}$: same data as holotype (cAss); $3\delta\delta$, 7_{Q} $_{Q}$, same data, but leg. Wunderle (cWun).

D e s c r i p t i o n : Measurements (in mm) and ratios (range, arithmetic mean; n=14): AL: 0.74-0.83, 0.78; HW: 0.35-0.41, 0.38; PL: 0.33-0.41, 0.38; PW: 0.40-0.47, 0.44; EL: 0.21-0.24, 0.23; HTi: 0.38-0.42, 0.40; HTa: 0.27-0.32, 0.29; ML: 0.39-0.41, 0.40; TL: 2.4-3.2, 2.8; PW/HW: 1.07-1.19, 1.14; PW/PL: 1.11-1.20, 1.15; EL/PL: 0.56-0.66, 0.61; HTa/HTi: 0.69-0.81, 0.73.

Species of intermediate size (see measurements) and dark colour. Whole body black, with the apex and the tergal margins only indistinctly lighter; antennae blackish brown; legs dark brown.

Head subcircular, approximately as long as wide; eyes relatively large and weakly projecting from lateral outline of head; postgenae in dorsal view ca. twice as long as eyes; puncturation shallow, but rather distinct; integument with microsculpture predominantly composed of transverse and diagonal striae and with weak shine. Antenna rather massive, distinctly incrassate apically; antennomeres I-III oblong, I and II of equal length, III distinctly shorter than II, IV-IX increasingly transverse, X less transverse (ca. 1.5x as wide as long) and longer than IX, and XI shorter than the combined length of IX and X.

Pronotum distinctly convex in cross-section, weakly transverse and slightly wider than head (see ratios PW/HW and PW/PL); maximal width at some distance from anterior angles; lateral margins in dorsal view weakly convex; posterior angles not very marked; midline usually more or less shallowly and narrowly impressed; puncturation finer and less distinct, microsculpture more distinct than on head, composed of irregular meshes and striae; surface with weaker shine than head; pubescence directed caudad in midline and more or less transversely laterad in lateral area.

Elytra shorter (see ratio EL/PL) and slightly wider than pronotum, weakly dilated posteriorly; anterior angles obliquely rounded, posterior margin obliquely truncate, near posterior

rior angles with moderate concavity; puncturation finely granulose and more distinct than that of pronotum; microsculpture predominantly composed of more or less transverse or diagonal meshes; surface slightly more shining than that of pronotum; pubescence weakly diagonal, directed more caudad than laterad. Hind wings reduced. Tibiae and tarsi relatively long (see measurements and ratio HTa/HTi); first metatarsomere longer than the combined length of the two following tarsomeres, sometimes almost as long as the combined length of the three following tarsomeres.

Abdomen weakly dilated caudad, maximal width across segment VI; terga III-V with distinct, tergum VI with weaker anterior impression; puncturation very finely granulose and relatively sparse; integument with microsculpture composed of transverse meshes and with some shine.

- 3: tergum VIII posteriorly weakly convex (Fig. 65); posterior margin of sternum VIII in the middle strongly convex, not distinctly pointed, and with long setae (Fig. 66); median lobe of aedeagus and apical lobe of paramere as in Figs. 61-64.
- Q: tergum VIII similar to that in \mathcal{S} , but wider (Fig. 67); sternum VIII posteriorly convex (Fig. 68); spermatheca as in Fig. 69.

D e r i v a t i o n o m i n i s: The name is derived from the type locality of the species, the Vrontous Oros in northeastern Greece.

C o m p a r a t i v e n o t e s: From all the preceding congeners, T. vrontousensis is readily distinguished by the sexual characters and by its dark colour. T. timfristosensis, which is nearly as dark, is much smaller and has light brown appendages. T. rastrifer and T. longiuter, which have a similar pronotal pubescence pattern, are larger and of lighter colour.

Distribution and bionomics: The types were collected on the western slope of the peak of the Vrontous Oros, Séres (=Sérrai), where they were sifted from grass, moss, and *Pinus* litter at an altitude of 1800m.

Tectusa sp. n.

In the Oros Vitsi, 3 $\circ \circ$ of a new species of *Tectusa* were collected together with *Pareudectus vitsiensis* sp. n. and a new species of *Coryphiodes* (Tab. 1). In external characters, this species is similar to *T. vrontousensis* sp. n. A description, however, is here refrained from because no $\circ \circ \circ$ are available and because it is uncertain if further similar species occur in the region.

Acknowledgements

We would like to express our thanks to the following specialists for their assistance in identifying Staphylinidae of certain genera: Johannes Frisch, Giessen (*Scopaeus* spp.), György Makranczy, currently Kansas (*Ochthephilus* spp.), Michael Schülke, Berlin (*Sepedophilus* spp.), Heinrich Terlutter, Billerbeck (*Ocalea* spp.), Jürgen Vogel, Görlitz (some *Atheta* spp.), and Adriano Zanetti, Verona (*Omalium* spp.). In addition, we are grateful to Harald Schillhammer, Naturhistorisches Museum Wien, for the loan of type material under his care and to Michael Schülke for his helpful comments on the manuscript.

Zusammenfassung

Die Ergebnisse einer Exkursion nach Zentral- und Nordgriechenland im Frühjahr 2000 werden vorgelegt. Von mehr als 214 Staphylinidenarten wurden 188 identifiziert, darunter eine Reihe von neuen Taxa und Erstnachweisen für Griechenland. 10 Arten werden beschrieben und von ähnlichen Arten unterschieden; wesentliche Differentialmerkmale werden abgebildet: Deliphrosoma angulatum sp. n. (Fokis, Fthiotis), Pareudectus vitsiensis sp. n. (Florina), Sunius fokisensis sp. n. (Fokis), Lathrobium vorasensis sp. n. (Florina), Tectusa rastrifera sp. n. (Fokis, Fthiotis), T. longiuter sp. n. (Evritania), T. viduus sp. n. (Evritania), T. timfristosensis sp. n. (Evritania), T. vardousiensis sp. n. (Fokis) und T. vrontousensis sp. n. (Séres). Anthobium ganglbaueri (LUZE), Quedius bernhaueri RAMBOUSEK, Leptusa jelineki PACE, L. peristerica PACE, L. winneguthiana PACE und L. sororella PACE werden erstmals aus Griechenland nachgewiesen. Der Habitus und der Aedoeagus von Q. bernhaueri werden abgebildet.

References

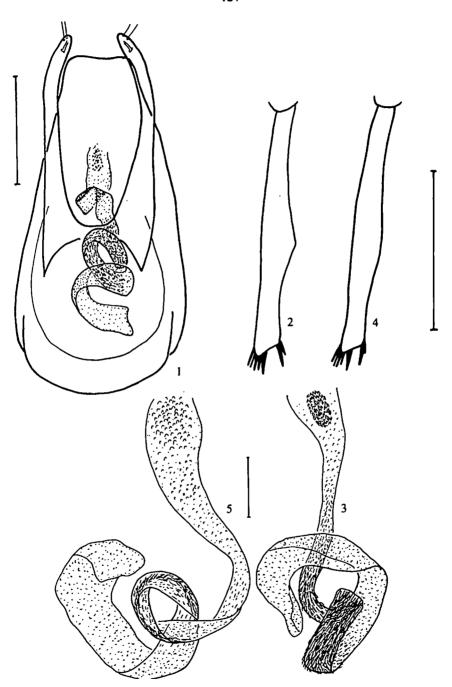
- ASSING V. (1995): Über Sunius fallax (LOKAY, 1919) (Col., Staphylinidae). Entomol. Nachr. Ber. 38 (1994): 267-269.
- ASSING V. (1997): A revision of *Othius* STEPHENS, 1829. III. The species of the Western Palaearctic region exclusive of the Atlantic Islands (Coleoptera, Staphylinidae: Xantholininae). Nova Suppl. Entomol., Berlin 10: 3-130.
- ASSING V. (1999): A revision of the species of *Geostiba* THOMSON 1958 from Greece and Cyprus (Coleoptera, Staphylinidae, Aleocharinae). Linzer biol. Beitr. 31: 849-928.
- ASSING V. (2000): A revision of the species of *Geostiba* THOMSON 1858 and *Paraleptusa* PEYERIMHOFF 1901 of Greece: Supplement I, including some species from Albania, Macedonia, Bulgaria, and Turkey (Coleoptera: Staphylinidae, Aleocharinae) Linzer biol. Beitr. 32/2: 1007-1031.
- ASSING V. & M. SCHÜLKE (2000): A new microphthalmous species of *Lathrobium* GRAVENHORST from Greece (Insecta: Coleoptera: Staphylinidae: Paederinae). Reichenbachia 32: 321-326.
- ASSING V. & P. WUNDERLE (1999): On the staphylinid fauna of northeastern Greece (Coleoptera). Entomol. Z. 109: 474-483.
- COIFFAIT H. (1972): Coléoptères Staphylinidae de la région paléarctique occidentale. I. Généralités; sous-fam. Xantholininae et Leptotyphlinae. Nouv. Rev. Ent. Suppl. 2 (2): 1-651.
- COIFFAIT H. (1974): Coléoptères Staphylinidae de la région paléarctique occidentale. II. Sous famille Staphilininae [sic] Tribus Philonthini et Stapyhlinini. Nouv. Rev. Ent. Suppl. 4 (4): 1-593.
- COIFFAIT H. (1978): Coléoptères Stapyhlinidae de la région paléarctique occidentale III. Sous famille Staphylininae, Tribu Quediini. Sous famille Paederinae, Tribu Pinophilini. Nouv. Rev. Ent. Suppl. 8 (4): 1-364.
- COIFFAIT H. (1982): Coléoptères Stapyhlinidae de la région paléarctique occidentale. IV. Sous famille Paederinae Tribu Paederini 1 (Paederi, Lathrobii). Nouv. Rev. Ent. Suppl. 12 (4): 1-440.
- COIFFAIT H. (1984): Coléoptères Stapyhlinidae de la région paléarctique occidentale. V. Sous famille Paederinae Tribu Paederini 2, Sous famille Euaesthetinae. Nouv. Rev. Ent. Suppl. 13 (4): 1-424.
- EPPELSHEIM E. (1888): IV. Staphylinen. Deutsche Ent. Zeitschr. (1888): 401-410.
- FAGEL G. (1968): Contribution a la connaissance des Staphylinidae. CII. Espèces inédites d'Anatolie et du bassin méditerranéen. Bull. Ann. Soc. R. Ent. Belg. 104: 118-134.

- Kraatz G. (1858a): Beitrag zur Käferfauna Griechenlands. Zweites Stück: Palpicornia, Silphales, Scydmaenidae, Pselaphidae, Staphylinidae. Berliner Ent. Zeitschr. 2: 37-67.
- KRAATZ G. (1858b): Beitrag zur K\u00e4ferfauna Griechenlands. Drittes St\u00fcck: Staphylinidae (Schluss), Trichopterygia, Histeridae, Phalacridae, Nitidulariae, Trogositarii, Colydii, Cucujidae, Cryptophagidae, Thorictidae, Mycetophagidae, Dermistini, Byrrhii. Berliner Ent. Zeitschr. 2: 123-148.
- LUZE G. (1905): Revision der paläarktischen Arten der Staphyliniden-Gattung Lathrimaeum Er. Verh. zool.-bot. Ges. Wien 55: 53-69.
- PACE R. (1984): Due nuovi Lathrobium del Durmitor e della Serbia (Coleoptera, Staphylinidae). In: Nonveiller G. (ed.): Prilozi Endogejskoj Fauni Tvrdokrilaca Durmitora (Insecta, Coleoptera). Fauna Durmitora Sveska 1: 349-359.
- PACE R. (1989): Monografia del genere Leptusa KRAATZ (Coleoptera, Staphylinidae). Mem. Mus. Civ. Stor. Nat. Verona (11° Ser.), Sez. Sci. Vita (A: Biol.) 8: 1-307.
- PUTHZ V. (1971): Zur Staphylinidenfauna des Balkans: Die bisher aus Jugoslawien und angrenzenden Ländern bekannten Steninen (Coleoptera, Staphylinidae). Wiss. Mitt. Bosnisch-herzegov. Landesmus. 1 (Heft C): 239-292.
- RAMBOUSEK F. G. (1915): Ein neues Subgenus der Gattung Quedius. Coleopt. Rdsch. 4: 130-132.
- SCHEERPELTZ O. (1931): XV. Teil. Staphylinidae (Coleoptera). In: BEIER M.: Zoologische Forschungsreise nach den Jonischen Inseln und dem Peloponnes. Sitzungsber. Akad. Wiss. Wien, Mathem.-naturw. Kl., Abt. I, 140: 359-460 + I-III.
- SCHEERPELTZ O. (1956): Die von Herrn Dr. E. Haaf vom Museum Frey-Tutzing gelegentlich seiner Studienreise 1955 in Griechenland und auf der Insel Cypern aufgesammelten Staphylinidae (Col.). Entomol. Arb. Mus. Frey 7: 1092-1117.
- SCHEERPELTZ O. (1957): Die von Herrn Fr. Borchmann gelegentlich seiner 1956 nach Griechenland unternommenen Studienreise aufgesammelten Staphyliniden (Col.). Entomol. Blätter 53: 151-155.
- SCHEERPELTZ O. (1959): VIII. Staphylinidae (Col.). In: BEIER M.: Zoologische Studien in West-Griechenland. Sitzungsber. Österr. Akad. Wiss., Mathem.-naturw. Kl., Abt. I, 167: 363-432.
- Scheerpeltz O. (1961): Neue Arten der Gattung Lathrimaeum Er., nebst einer Bestimmungstabelle der bisher bekannt gewordenen palaearktischen Arten dieser Gattung (Col. Staphylinidae). Mitt. Münchn. Ent. Ges. 51: 72-95.
- SCHEERPELTZ O. (1963): Wissenschaftliche Ergebnisse eines Sammelaufenthaltes Prof. Dr. K. Mandl's auf der Insel Rhodos (Col. Staph.). Koleopt. Rdsch. 40/41 (1962/63): 67-76.
- SCHEERPELTZ O. (1964): Staphyliniden von der Insel Kreta (Coleoptera). Ann. Hist.-Nat. Mus. Nat. Hung. 56: 297-308.
- ZERCHE L. (1990): Monographie der paläarktischen Coryphiini (Coleoptera, Staphylinidae, Omaliinae). Berlin: Akad. d. Landwirtschaftswiss. d. DDR: 1-413.
- ZERCHE L. (1991): Beitrag zur Taxonomie und Verbreitung der Gattung *Deliphrosoma* REITTER, 1909 (Coleoptera, Staphylinidae, Omaliinae). Beitr. Ent. Berlin 41: 313-332.
- ZERCHE L. (1998): Neue *Deliphrosoma*-Arten aus Bulgarien und aus der Türkei und neue Unterarten von *Deliphrosoma prolongatum* (ROTTENBERG) aus Bulgarien (Insecta: Coleoptera: Staphylinidae: Omaliinae). Reichenbachia 32: 245-255.

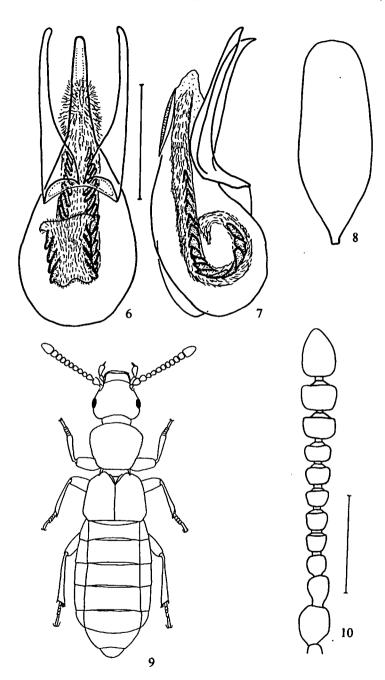
Authors' addresses:

Volker ASSING Gabelsbergerstr. 2 D-30163 Hannover, Germany e-mail: vassing.hann@t-online.de

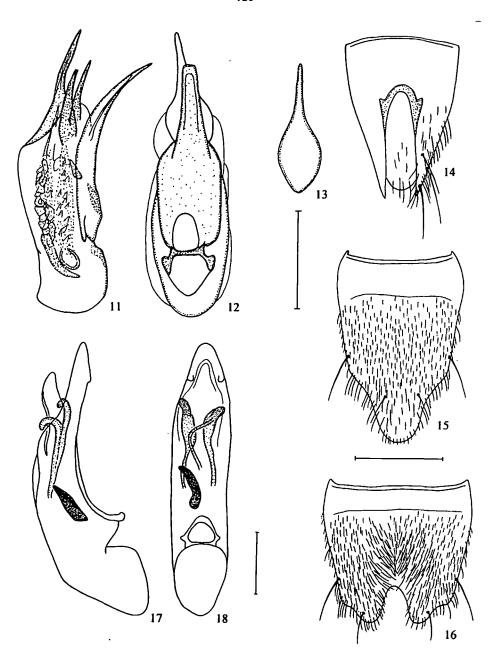
Paul WUNDERLE Hehnerstr. 15 41069 Mönchengladbach, Germany e-mail: p.wunderle.mg@t-online.de



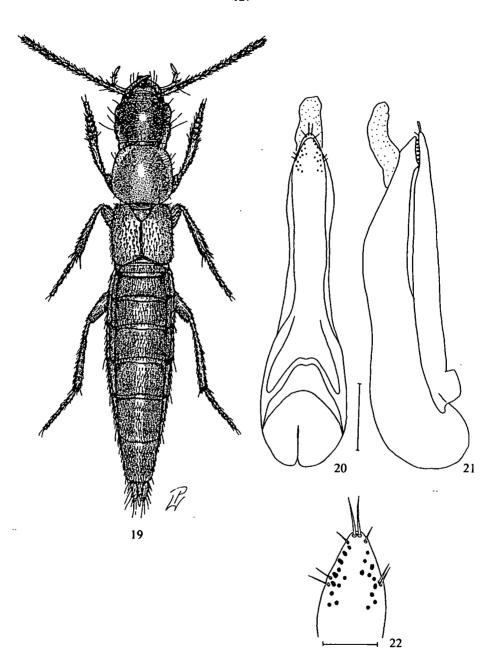
Figs. 1-5: Deliphrosoma angulatum sp. n. (1-3) and Deliphrosoma fratellum (ROTTENBERG) (4-5): 1- aedeagus in ventral view (holotype); 2, $4-\delta$ protibia; 3, 5- internal tube of aedeagus. Scales: 1, 3, 5: 0.25 mm; 2, 4: 0.5 mm.



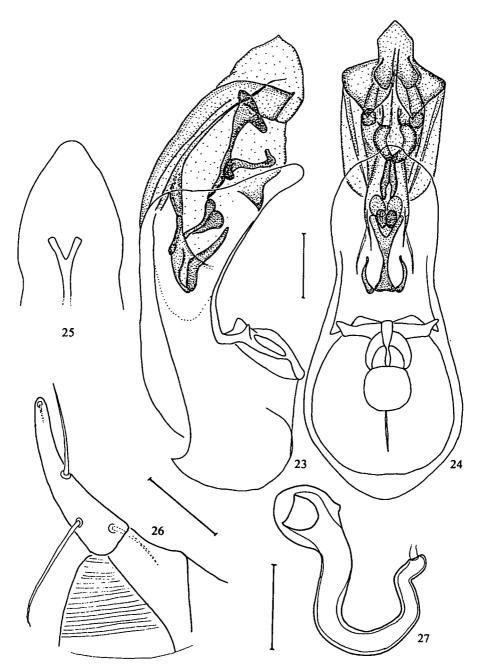
Figs. 6-10: Pareudectus vitsiensis sp. n.: 6, 7 –aedeagus (holotype) in ventral and in lateral view; 8 – \eth sternum IX; 9 – habitus of \lozenge (length: 2.75 mm); 10 – antenna. Scale: 6-8, 10: 0.25 mm; 9: without scale.



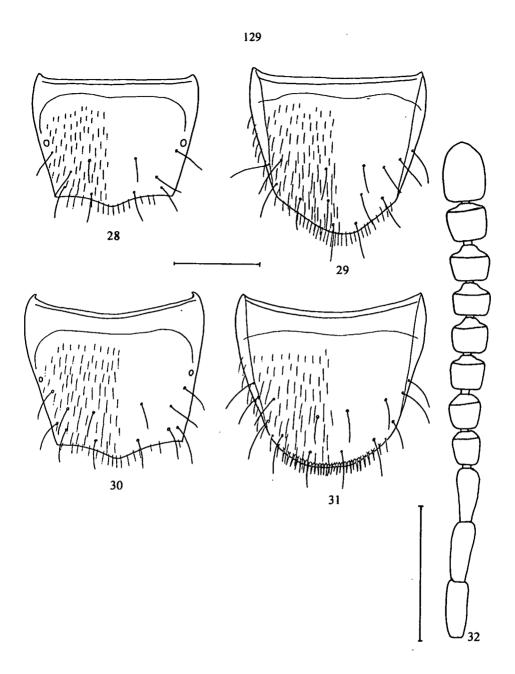
Figs. 11-18: Lathrobium vorasensis sp. n. (11-16) and Sunius fokisensis sp. n. (17-18): 11, 12, 17, 18 – aedeagus in lateral and in ventral view; 13 – dorso-apical sclerite of aedeagus; 14 - 0 abdominal segments IX and X in dorsal view; 15 - 0 sternum VIII; 16 - 0 sternum VIII. Scales: 11-16: 0.5 mm; 17-18: 0.1 mm.



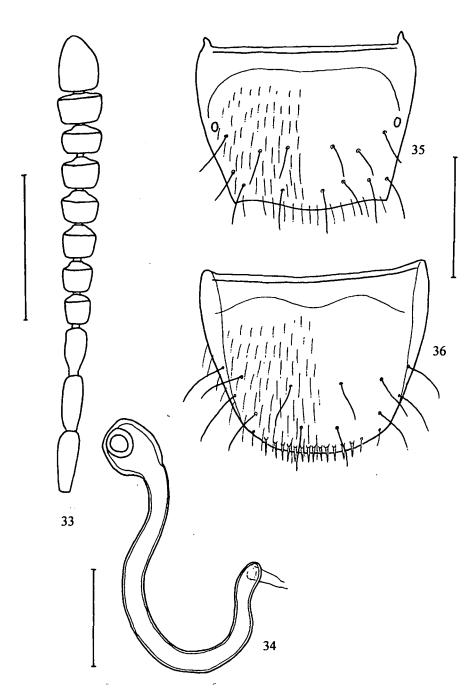
Figs. 19-22: Quedius bernhaueri RAMBOUSEK: 19 – facies (length: 11.0 mm); 20, 21 – aedeagus in ventral and in lateral view; 22 – apex of paramere. Scales: 20-21: 0.25 mm; 22: 0.1 mm; 19: without scale.



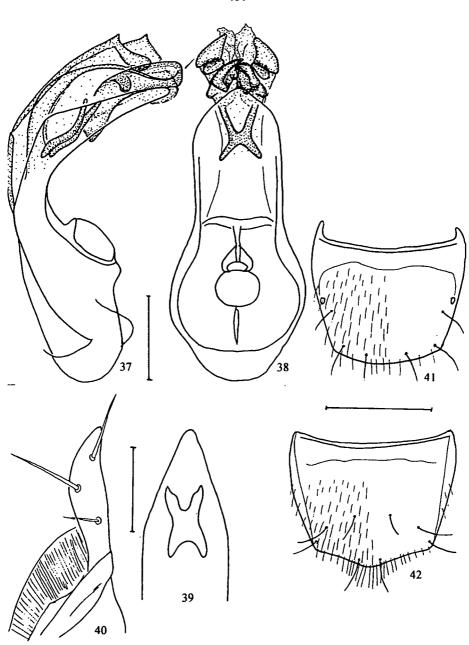
Figs. 23-27: *Tectusa rastrifera* sp. n.: 23, 24 – median lobe of aedeagus in lateral and in ventral view; 25 – apex of ventral process of median lobe in antero-ventral view; 26 – apex of paramere; 27 – spermatheca. Scales: 0.1 mm.



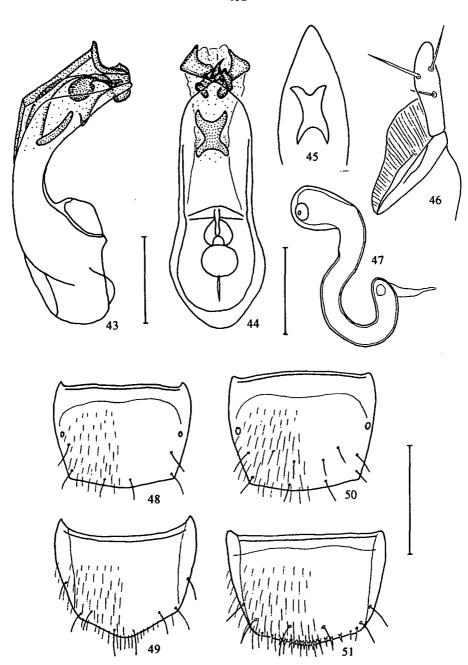
Figs. 28-32: Tectusa rastrifera sp. n.: 28 – δ tergum VIII; 29 – δ sternum VIII; 30 – φ tergum VIII; 31 – φ sternum VIII; 32 – antenna. Scales: 0.25 mm.



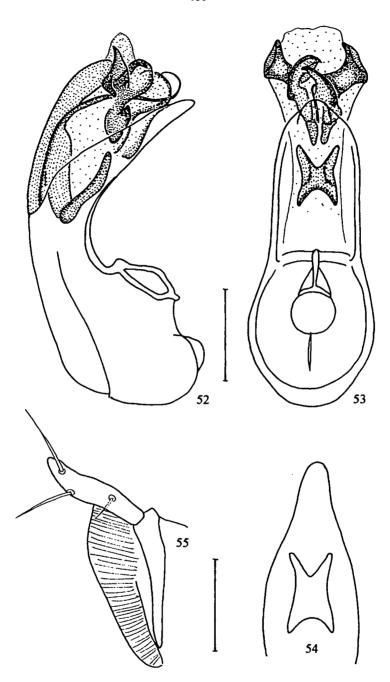
Figs. 33-36: Tectusa longiuter sp. n.: 33 – antenna; 34 – spermatheca; 35 – φ tergum VIII; 36 – φ sternum VIII. Scales: 34: 0.1 mm; 33, 35, 36: 0.25 mm.



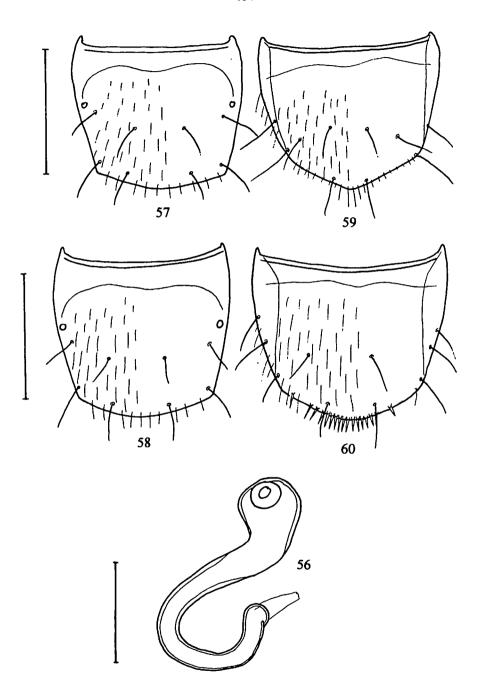
Figs. 37-42: Tectusa viduus sp. n.: 37, 38 – median lobe of aedeagus in lateral and in ventral view; 39 – apex of ventral process of median lobe in antero-ventral view; 40 – apex of paramere; 41 - 3 tergum VIII; 42 - 3 sternum VIII. Scales: 37-40: 0.1 mm; 41, 42: 0.25 mm.



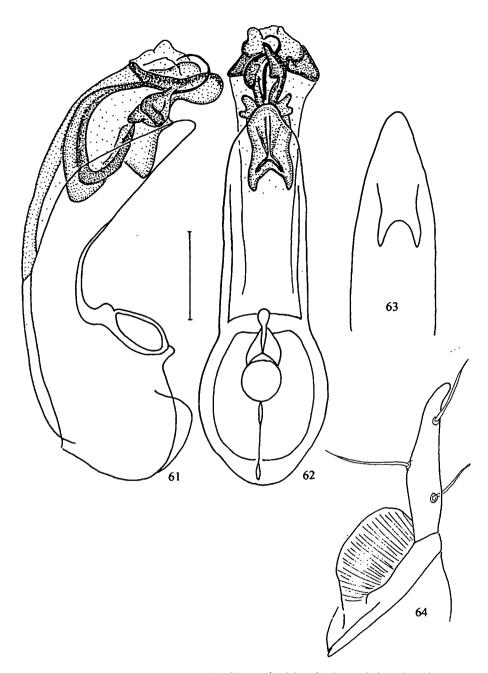
Figs. 43-51: Tectusa timfristosensis sp. n.: 43, 44 — median lobe of aedeagus in lateral and in ventral view; 45 — apex of ventral process of median lobe in antero-ventral view; 46 — apex of paramere; 47 — spermatheca; 48 — σ tergum VIII; 49 — σ sternum VIII; 50 — ρ tergum VIII; 51 — ρ sternum VIII. Scales: 43-47: 0.1 mm; 48-51: 0.25 mm.



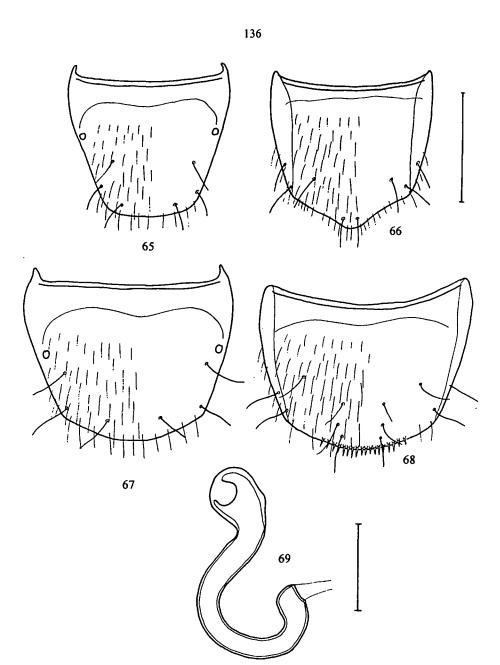
Figs. 52-55: Tectusa vardousiensis sp. n.: 52, 53 – median lobe of aedeagus in lateral and in ventral view; 54 – apex of ventral process of median lobe in antero-ventral view; 55 – apex of paramere. Scales: 0.1 mm.



Figs. 56-60: Tectusa vardousiensis sp. n.: 56 – spermatheca; 57 – \eth tergum VIII; 58 – ϱ tergum VIII; 59 – \eth sternum VIII; 60 – ϱ sternum VIII. Scales: 56: 0.1 mm; 57-60: 0.25 mm.



Figs. 61-64: *Tectusa vrontousensis* sp. n.: 61, 62 – median lobe of aedeagus in lateral and in ventral view; 63 – apex of ventral process of median lobe in antero-ventral view; 64 – apex of paramere. Scale: 0.1 mm.



Figs. 65-69: Tectusa vrontousensis sp. n.: 65 – \eth tergum VIII; 66 – \eth sternum VIII; 67 – \lozenge tergum VIII; 68 – \lozenge sternum VIII; 69 – spermatheca. Scales: 69: 0.1 mm; 65-68: 0.25 mm.